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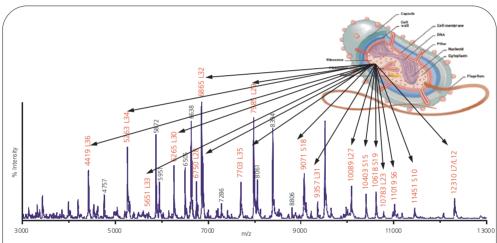


## 1. Introduction

Whole-Cell MALDI-TOF MS (WC-MS) is the most widely used mass-based approaches for bacterial identification and classification. Commercialized bacterial identification systems on the basis of protein mass fingerprinting are rapidly expanding. However, the mass fingerprinting methodology can not assure to discriminate bacteria at strain level, partly because strain specific biomarker based on bacterial genomic database has not been assigned with the method. Until now we have proposed a reliable

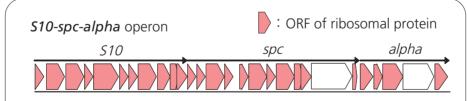
phylogenetic classification method at the strain level by MALDI-TOF MS using ribosomal subunit proteins coded in the *S10-spc-alpha* operon, which encodes half of the ribosomal proteins in eubacterial genomes, as proteomic biomarkers.

In this study, the method, named *S10*-GERMS (*S10-spc-alpha* operon Gene Encoded Ribosomal Protein Mass Spectrum) was applied to discriminate the pathovar of *P. syringae*, a plant-pathogenic bacterium.



- $\bullet$  ~50% of detected peaks in WC -MS (> m/z 4000) are derived from ribosomal proteins.
- Assigned ribosomal subunit proteins based on DNA sequences and WC-MS can be specific markers for bacterial classification at species and strain level <sup>1), 2), 3)</sup>.
- However, available DNA sequence information of ribosomal proteins is limited, because most of bacterial strains are not genome-sequenced.

Fig. 1 Typical example of WC-MS spectrum



- *S10-spc-alpha* operon contains more than ribosomal subunits, which are commonly conserved in bacteria.
- At the moment, DNA sequencing of this operon using conventional Sanger DNA sequencer is more accessible and accurate than whole genome sequencing.
- Using gene sequence of this operon, a considerable number of WC-MS peaks of bacterial strain can be assigned even if the strain is not genome sequenced.

Fig. 2 Diagram of S10-spc-alpha operon



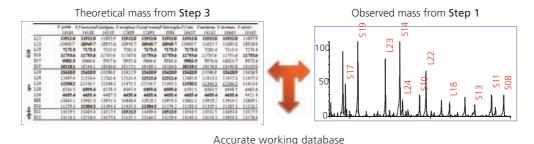
# 2. Methods and Materials

Workflow of the phylogenetic biomarker finding and constructing database

- Step 1- Analyzing bacterial strains by WC-MS (observed mass).
- Step 2- DNA sequencing and translating of S10-spc-alpha operon of the strains.
- **Step 3-** Construction of theoretical ribosomal proteins database of the strains.

  Theoretical mass of each subunit protein was calculated taking account of N-terminal methionine loss.
- **Step 4** Construction of accurate database by comparison of theoretical vs. observed mass.

  To validate the constructed database, the theoretical masses of MS analyzed bacterial strains at step 1 were compared with each of the MALDI mass spectra.



P. fluorescens P.azotoformans P.chlororaphis P. aeruginosa P.mendocina 11893 91 L23 10900.65 10900.65 10950.71 11015.74 11085.83 10920.64 10955.60 L 29 7173.31 7173.31 7173.31 7173.31 7202.35 7205.35 7215.39 11753.58 11753.58 11783.61 11783.61 11753.58 11753.58 11753.58 11767.61 11755.55 9955.60 S 19 10246 12 1018907 1020404 10175 98 10190.01 L 18 12556,43 12413.29 12457 30 12485.36 L24 11336.25 11471 46 11336.25 11344.27 11344.27 11413.37 11340 32 11345.26 L30 6347.44 6292.46 6363.48 L36 S08 13920.23 13869.10 13973.29 14040.42 13928.16 13914.18 13951.25 S 14 11274.24 13210.43 13164.45

Using this database, discrimination of sample bacteria at the strain level was performed.

#### **Bacterial Strains**

Commercially available strains of *Pseudomonas syringae* were purchased from the National Institute of Technology and Evaluation (NITE)-Biological Resource Center (NBRC, Kisarazu, Japan)

*P. syringae* NBRC 3310, NBRC 3508, NBRC 12655, NBRC 12656, NBRC 14053, NBRC 14083 and NBRC 14084. Each bacterial strain was grown aerobically in the medium and at the temperature recommended by its suppliers.

#### Construction of the ribosomal protein database

The amino acid sequences of all ribosomal subunit proteins of the *P. syringae* strains were obtained from the NCBInr database. The theoretical mass of each subunit protein as [M + H]\* was calculated from translated amino acid sequences using a Compute pl/Mw tool on the ExPASy proteomics server. N-Terminal methionine loss was only considered based on the 'N-end rule' as a possible

post-translational modification. When there were some errors in their theoretical masses they were corrected by comparison with the observed masses by WC-MS. The selected biomarker proteins based on the corrected ribosomal protein database were used for bacterial discrimination at the strain level of *P. syringae*.



#### **MALDI-TOFMS**

Instrument : AXIMA Performance (Shimadzu Biotech/Kratos)

Tuning mode : Linear positive

Matrix solution : Sinapinic acid 10 mg/mL (50% acetonitrile 1% TFA)

Sample preparation: Bacterial cells were harvested by centrifugation and washed twice in TMA-I buffer (10 mM Tris-HCI

(pH 7.8), 30 mM NH<sub>4</sub>Cl, 10 mM MgCl<sub>2</sub>, and 6 mM 2-mercaptoethanol). Each harvested solution of whole cells was mixed with the matrix solution. Then, the mixed solution was spotted onto the

MALDI target and dried in air.

## 3. Results

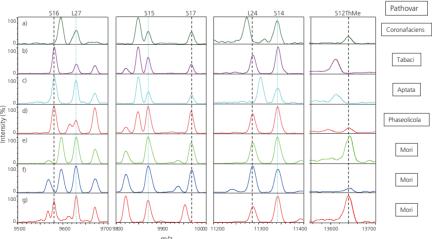
In this study, the *S10*-GERMS method was applied for discrimination of the pathovar of *P. syringae* using genome-sequenced and commercially available strains as model samples. Theoretical masses of ribosomal proteins were calculated by *S10-spc-alpha* operon sequence information of the 12 genome-sequenced strains of *P. syringae*. 14 ribosomal subunit proteins with less than *m/z* 

15,000, coded in the *S10-spc-alpha* operon (except for L14) and two additional ribosomal proteins (S16, S12) were selected as reliable and reproducible biomarkers for typing the pathovar of *P. syringae* (Table 1). Several peaks seem to be pathovar-specific. And then, 7 sample strains of *P. syringae* were analyzed by MALDI-TOF MS (Fig. 3).

Table 1 Theoretical mass of genome-sequenced strains of P. syringae

| Protein<br>name | Coded — |         | Theoretical masses as [M + H]* |              |         |          |          |         |         |              |         |         |         |  |  |  |
|-----------------|---------|---------|--------------------------------|--------------|---------|----------|----------|---------|---------|--------------|---------|---------|---------|--|--|--|
|                 |         | Aesculi | Aesculi                        | Phaseolicola | Tabaci  | Syringae | Syringae | Oryzae  | Tomato  | Tomato       | Tomato  | Tomato  | Tomato  |  |  |  |
|                 | operon  | 2250    | NCPPB 3681                     | 1448A ATCC 1 | 1528    | B728a    | 642      | 1_6     | K40     | Max13NCPPB 1 | 108     | DC3000  | T1      |  |  |  |
| L36             | spc     | 4435.4  | 4435.4                         | 4435.4       | 4435.4  | 4435.4   | 4435.4   | 4435.4  | 4435.4  | 4435.4       | 4435.4  | 4435.4  | 4435.4  |  |  |  |
| L30             | spc     | 6422.7  | 6422.7                         | 6422.7       | 6422.7  | 6422.7   | 6408.6   | 6422.7  | 6422.7  | 6422.7       | 6422.7  | 6422.7  | 6426.7  |  |  |  |
| L29             | S10     | 7173.3  | 7173.3                         | 7173.3       | 7173.3  | 7173.3   | 7173.3   | 7173.3  | 7173.3  | 7173.3       | 7173.3  | 7173.3  | 7173.3  |  |  |  |
| S17             | S10     | 9966.6  | 9966.6                         | 9966.6       | 9966.6  | 9966.6   | 9966.6   | 9966.6  | 9966.6  | 9966.6       | 9966.6  | 9966.6  | 9966.6  |  |  |  |
| S19             | S10     | 10204.0 | 10204.0                        | 10204.0      | 10204.0 | 10204.0  | 10204.0  | 10204.0 | 10177.0 | 10177.0      | 10177.0 | 10177.0 | 10177.0 |  |  |  |
| L23             | S10     | 10945.7 | 10945.7                        | 10945.7      | 10945.7 | 10945.7  | 10945.7  | 10945.7 | 10945.7 | 10945.7      | 10945.7 | 10945.7 | 10945.7 |  |  |  |
| L24             | spc     | 11288.2 | 11288.2                        | 11288.2      | 11288.2 | 11306.2  | 11306.2  | 11274.2 | 11288.2 | 11288.2      | 11288.2 | 11288.2 | 11288.2 |  |  |  |
| S14             | spc     | 11362.4 | 11362.4                        | 11343.3      | 11343.3 | 11346.4  | 11343.3  | 11343.3 | 11343.3 | 11343.3      | 11343.3 | 11343.3 | 11343.3 |  |  |  |
| S10             | S10     | 11796.6 | 11796.6                        | 14733.2      | 11767.6 | 11767.6  | 11767.6  | 11767.6 | 11767.6 | 11767.6      | 11767.6 | 11767.6 | 11767.6 |  |  |  |
| L22             | S10     | 11893.9 | 11893.9                        | 11893.9      | 11893.9 | 11893.9  | 11893.9  | 11893.9 | 11893.9 | 11893.9      | 11893.9 | 11893.9 | 11893.9 |  |  |  |
| L18             | spc     | 12473.4 | 12473.4                        | 12473.4      | 12473.4 | 12473.4  | 12473.4  | 12473.4 | 12473.4 | 12473.4      | 12473.4 | 12473.4 | - 💥     |  |  |  |
| S13             | alpha   | 13239.5 | 13239.5                        | 13239.5      | 13239.5 | 13239.5  | 13239.5  | 13239.5 | 13239.5 | 13239.5      | 13239.5 | 13239.5 | 13239.5 |  |  |  |
| S11             | alpha   | 13485.4 | 13485.4                        | 13485.4      | 13485.4 | 13485.4  | 13485.4  | 13485.4 | 13485.4 | 13485.4      | 13485.4 | 13485.4 | 13485.4 |  |  |  |
| S08             | spc     | 13892.1 | 13892.1                        | 13892.1      | 13892.1 | 13892.1  | 13892.1  | 13892.1 | 13892.1 | 13892.1      | 13892.1 | 13892.1 | 13892.1 |  |  |  |
| S16             |         | 9580.9  | 9580.9                         | 9580.9       | 9580.9  | 9580.9   | 9580.9   | - *     | 9580.8  | 9594.9       | 9594.9  | 9594.9  | 9594.9  |  |  |  |
| S12             |         | 13594.7 | 13594.7                        | 13594.7      | 13594.7 | 13594.7  | 13594.7  | 13594.7 | 13628.7 | 13594.7      | 13594.7 | 13594.7 | 13594.7 |  |  |  |

\* There was a mis-sequence in the genome sequence



P. syringae: NBRC 3310 (a), NBRC 3508 (b), NBRC 12655 (c), NBRC 12656 (d), NBRC 14053 (e), NBRC 14083 (f), and NBRC 14084 (g).

Fig. 3 MALDI mass spectra of *P. syringae*: NBRC 3310 (a), NBRC 3508 (b), NBRC 12655 (c), NBRC 12656 (d), NBRC 14053 (e), NBRC 14083 (f), and NBRC 14084 (g).



In this analysis, 2 ribosomal subunits, L24 and S17 which are encoded in the *S10-spc-alpha* operon, were biomarkers for discrimination of these strains. Furthermore, MALDI-TOF MS analysis revealed that 2 ribosomal subunits, S12 and S16, play a useful role in discrimination of the strain of *P. syringae*.

Using these ribosormal protein biomarkers, the 7 sample strains were classified into 5 clusters as color-coded (Table 2). Each cluster seems to reflect their pathovar. Further analysis of other *P. syringae* strains is needed to confirm this point.

Table 2 Ribosomal protein profiling table of *P. syringae* strains.

| D-th                  | Strain -               | Ribosomal protein types |     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |
|-----------------------|------------------------|-------------------------|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Pathovar              |                        | L36                     | L30 | L29 | S17 | S19 | L23 | L24 | S14 | S10 | L22 | L18 | S13 | S11 | S08 | S16 | S12 |
| Genomesequ            |                        |                         |     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |
| Aesculi               | 2250                   | 1                       | I   | - 1 | - 1 | - 1 | - 1 | 1   | 1   | - 1 | - 1 | 1   | - 1 | - 1 | 1   | 1   | -   |
| Aesculi               | NCPPB 3681             | 1                       | I   | - 1 | - 1 | - 1 | - 1 | 1   | 1   | - 1 | - 1 | 1   | - 1 | - 1 | 1   | 1   | -   |
| Phaseolicola          | 1448A                  | 1                       | I   | - 1 | - 1 | - 1 | - 1 | 1   | II  | Ш   | - 1 | 1   | - 1 | - 1 | 1   | 1   | -   |
| Tabaci                | ATCC 11528             | I                       | I   | I   | I   | 1   | I   | I   | Ш   | П   | 1   | 1   | I   | I   | 1   | 1   |     |
| Syringae              | B728a                  | I                       | I   | I   | I   | 1   | I   | Ш   | Ш   | П   | I   | 1   | I   | I   | I   | I   | - 1 |
| Syringae              | 642                    | I                       | II  | 1   | I   | 1   | I   | Ш   | Ш   | П   | I   | I   | I   | I   | 1   | 1   |     |
| Oryzae                | 1_6                    | I                       | I   | I   | I   | 1   | I   | Ш   | Ш   | П   | I   | 1   | -   | I   | I   | -   | - 1 |
| Tomato                | K40                    | 1                       | I   | I   | 1   | Ш   | 1   | 1   | Ш   | П   | 1   | 1   | I   | 1   | 1   | 1   | П   |
| Tomato                | Max13                  | I                       | I   | I   | 1   | П   | 1   | 1   | Ш   | П   | 1   | 1   | 1   | 1   | 1   | II  |     |
| Tomato                | NCPPB 1108             | 1                       | I   | I   | 1   | II  | 1   | 1   | Ш   | П   | 1   | 1   | 1   | 1   | 1   | II  | 1   |
| Tomato                | DC3000                 | I                       | I   | I   | 1   | Ш   | 1   | 1   | II  | II  | 1   | 1   | I   | 1   | 1   | II  | 1   |
| Tomato                | T1                     | 1                       | Ш   | I   | 1   | II  | 1   | 1   | Ш   | П   | 1   | -   | 1   | 1   | 1   | II  | 1   |
| Sample strain         | IS                     |                         |     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |
| CoronafaciensNBRC3310 |                        |                         | I   | I   |     |     |     | Ш   | Ш   |     | I   | I   |     | -   | I   | Ш   |     |
| Tabaci                | NBRC3508               |                         |     |     |     |     |     |     | Ш   |     |     |     |     |     |     |     |     |
| Aptata                | NBRC12655              |                         | I   | I   |     | I   | I   | Ш   | Ш   | I   |     | I   |     | I   |     | I   |     |
| Phaseolicola          | Phaseolicola NBRC12656 |                         |     |     |     |     |     |     | Ш   |     |     |     |     |     | I   | I   |     |
| Mori                  | NBRC14053              | I                       | I   | I   | I   | I   | I   | I   | Ш   | I   | I   | I   | I   | I   | - [ | П   | Ш   |
| Mori                  | NBRC14083              |                         |     |     |     |     |     |     | Ш   |     |     |     |     |     |     | Ш   | Ш   |
| Mori                  | NBRC14084              | I                       | I   | ı   | Ш   |     |     |     | П   |     | I   | I   |     |     | I   | Ш   |     |

<sup>1 :</sup>Type of the ribosomal proteins of *P. syringae* 2250 strain.

# 4. Conclusions

- The *S10*-GERMS (*S10-spc-alpha* operon gene encoded ribosomal protein mass spectrum) method is a significantly useful tool for bacterial discrimination at strain level because WC-MS is significantly simpler and faster than DNA-based classification methods.
- Using gene sequence of S10-spc-alpha operon, a considerable number of WC-MS peaks of bacterial strain can be assigned even if the strain is not genome sequenced.
- This *S10*-GERMS and relevant method may have potential for the rapid discrimination of *P. syringae* at the pathovar as well as the strain level.
- The S10-GERMS method can expand the potential of the WC-MS into strain typing, which is important for bacterial outbreak analysis and tracking sources of bacterial contamination.

II and III: the mass of the biomarkers of other *P. syringae* strains which are differed from type I.



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