

IATDMCT 2017

Eishi IMOTO¹, Aurore JAFFUEL², Fanny DAYOT³, Jean-François HOEFFLER³, Alban HUTEAU²

¹SHIMADZU Corporation, Kyoto, Japan,

²SHIMADZU France, Noisiel, France,

³ALSACHIM,IIIkirch, France





Introduction

Measurement of immunosuppressant drugs (Figure 1.) is essential during organ transplantation. Under-dosing can lead to organ rejection, while over-dosing can cause serious toxicity. Traditional methods to measure immunosuppressant drugs in whole blood are based on either immunoassays or chromatography. Immunoassays, though, are affected by matrix interferences and lack of specificity. LC-MS/MS has then become the gold standard due to its specificity, precision and sensitivity. However, it

has still one major drawback: current LC-MS/MS platforms demand personnel with expertise and, for whole blood samples, tedious sample preparation. As a consequence, sample throughput is generally much lower than for immunoassays. We here report a fully automated procedure for the quantitation of four major immunosuppressant in whole blood samples, using of 13C labelled internal standards.

Figure 1: Structures of tacrolimus (a), sirolimus (b), everolimus (c), and cyclosporine-A (d)



Method and Materials

The quantitative analysis of Immunosuppressant (Figure 2.) was performed using reagents provided in Alsachim Dosimmune® kit. The Immunosuppressant and Internal standard were monitored using UHPLC-MS/MS system (Nexera X2 and LCMS-8050, Shimadzu, Kyoto). Sample preparation was performed using extraction buffer and

internal standard set provided in Alsachim Dosimmune® kit. Analytical performance of the method was monitored using whole blood calibrators and whole blood QC. Automatic sample preparation was performed using CLAM-2000 module (Shimadzu, Kyoto).

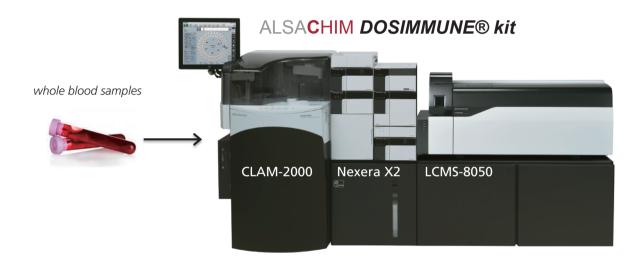


Figure 2: Sample workflow overview.

Sample preparation: CLAM-2000 and Dosimmune® kit

Whole blood sample tube is placed into the CLAM-2000 system: (1) 25μ L of whole blood sample is mixed with 12.5μ L of Internal standard and 175μ L of extraction buffer solution, (2) followed by a 30s stirring and (3) a 1min filtration. The extracted sample is transferred to the autosampler of the Nexera X2 system and injected immediately.

UHPLC conditions: Nexera X2 and Dosimmune® kit

Analytical column : Ascentis® C18 2,1x50 mm, 5 µm Trap column : Ascentis® C8 4,6x30 mm, 5 µm

Injection volume : 20 μL

Mobile Phase A : 90% 3mM Ammonium formate (pH=3.6) 10% MeOH Mobile Phase B : 10% 3mM Ammonium formate (pH=3.6) 90% MeOH

Isocratic flow rate : Mobile Phase A: 2 mL/min (trap column),

Mobile Phase B: 0.8 mL/min (analytical column)

Oven temperature: 65°C

MS conditions: LCMS-8050

Nebulizing Gas: 3 L/min (N2)HESI: 200°CPause time: 1 msecHeating Gas: 10 L/min (Air)DL: 250°CPolarity switching: 5 msecDrying Gas: 10 L/min (N2)HB: 200°CPoints per peak: > 30



Compound Formula Monoisotopic Mass MRM **Everolimus** C₅₃H₈₃NO₁₄ 957,6 975,6 → 908,5 Everolimus 13C2d4 963,6 $981,5 \rightarrow 914,5$ $C_{51}^{13}C_2H_{79}D_4NO_{14}$ 931,6 → 864,5 Sirolimus C₅₁H₇₉NO₁₃ 913,5 $935,4 \rightarrow 864,5$ Sirolimus 13Cd3 $C_{50}^{13}CH_{76}D_3NO_{13}$ 917,5 Tacrolimus C₄₄H₆₉NO₁₂ 803,5 $821,5 \rightarrow 768,6$ Tacrolimus 13Cd₄ $826,4 \rightarrow 773,6$ $C_{43}^{13}CH_{67}D_4NO_{12}$ 808.5 1219,9 → 1202,8 Ciclosporine $C_{62}H_{111}N_{11}O_{12}$ 1201,8 $1231.8 \rightarrow 1214.9$ Ciclosporine d₁₂ $C_{62}H_{99}D_{12}N_{11}O_{12}$ 1213.8

Table 1: Formula, exact mass and MRM transition for each compound

Results

Method conditions

The method enables the quantification of tacrolimus, sirolimus, everolimus and cyclosporine-A in whole blood samples. The established quantification strategy for this compounds is to use internal calibration using deuterium labeled standards. However, they generally suffer from poor isotopic enrichment, leading to overestimation of the unlabeled form. We here use 13C labeled internal standards for tacrolimus, sirolimus and everolimus. This guaranties better isotopic enrichment, better precision of

the results, long term stability of the standards and perfect co-elution with the analytes, leading to a better correction of matrix effects. Linearity was confirmed in the range 0.5 to 40 ng/mL for tacrolimus, sirolimus, everolimus, and in the range 5 to 1500 ng/mL for cyclosporine-A (Figure 3.). For all analytes, r² of linearity models was above 0.99, with S/N > 25 for LLOQ levels (Figure 4.). Controls showed accuracies comprised in between 85 and 115% for all analytes (Table 2.).

Calibration in whole blood

Linearity was confirmed, in whole blood, in the range 0.5-40 ng/mL for tacrolimus (Figure 3.a.), sirolimus (Figure 3.b.), everolimus (Figure 3.c.), and 5-1500 ng/mL for cyclosporine-A (Figure 3.d.).



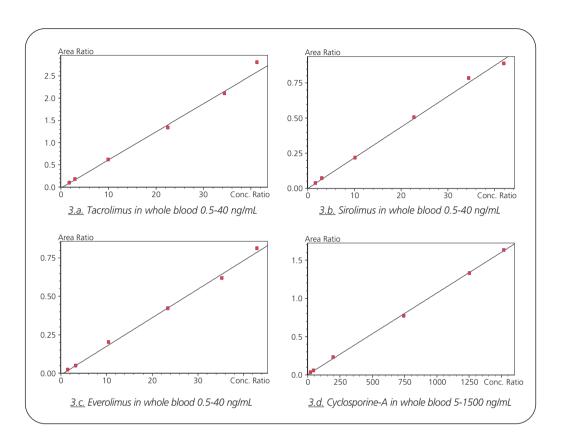


Figure 3: Calibration curves for tacrolimus (a), sirolimus (b), everolimus (c), and cyclosporine-A (d), in whole blood.

Limits of quantification in whole blood

The limits of quantification (LLOQ), in whole blood, are 0.5 ng/mL for tacrolimus, sirolimus, and everolimus, and 5 ng/mL for cyclosporine-A. The signal to noise ratio is above 25 at LLOQ levels.

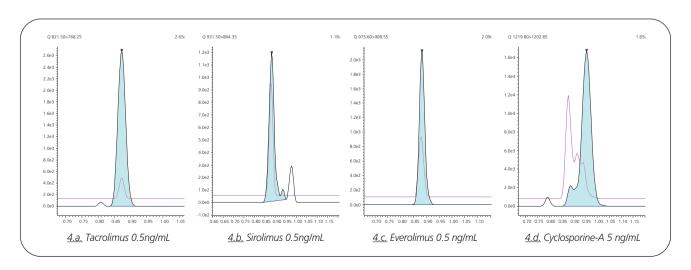


Figure 4: MRM chromatograms, at LLOQ levels, for tacrolimus (a), sirolimus (b), everolimus (c), and cyclosporine-A (d), in whole blood.



Performance Evaluation

Whole blood controls showed accuracies comprised in between 85% and 115% for all analytes.

Table 2: Whole blood control samples accuracies.

Tacrolimus	
Conc. (µg/mL)	Accuracy (%)
2.9	114
5.4	108
13.3	87
40.5	95

Sirolimus	
Conc. (µg/mL)	Accuracy (%)
3.2	115
5.7	105
13.3	93
40.6	103

Everolimus	
Conc. (µg/mL)	Accuracy (%)
3.2	114
5.8	92
13.4	85
42.1	94

Cyclosporine-A		
Conc. (µg/mL)	Accuracy (%)	
36.1	98	
223.4	106	
454.6	85	
1693	95	

Conclusion

Fully automated sensitive quantification of immunosuppressant drugs in whole blood, using high quality 13C labelled internal standards, increasing data quality, throughput and safety.

For Research Use Only. Not for use in diagnostic procedures. Not available in the USA, Canada, and China.



Shimadzu Corporation www.shimadzu.com/an/

For Research Use Only. Not for use in diagnostic procedures.

This publication may contain references to products that are not available in your country. Please contact us to check the availability of these products in your country.

The content of this publication shall not be reproduced, altered or sold for any commercial purpose without the written approval of Shimadzu. Company names, products/service names and logos used in this publication are trademarks and trade names of Shimadzu Corporation, its subsidiaries or its affiliates, whether or not they are used with trademark symbol "TM" or "®".

Third-party trademarks and trade names may be used in this publication to refer to either the entities or their products/services, whether or not they are used with trademark symbol "TM" or "®". Shimadzu disclaims any proprietary interest in trademarks and trade names other than its own.

The information contained herein is provided to you "as is" without warranty of any kind including without limitation warranties as to its accuracy or completeness. Shimadzu does not assume any responsibility or liability for any damage, whether direct or indirect, relating to the use of this publication. This publication is based upon the information available to Shimadzu on or before the date of publication, and subject to change without notice.

First Edition: November, 2017